

CLONING AND CHARACTERIZATION OF HERBICIDE-DEGRADING GLUTATHIONE TRANSFERASES FROM *CICER ARIETINUM*

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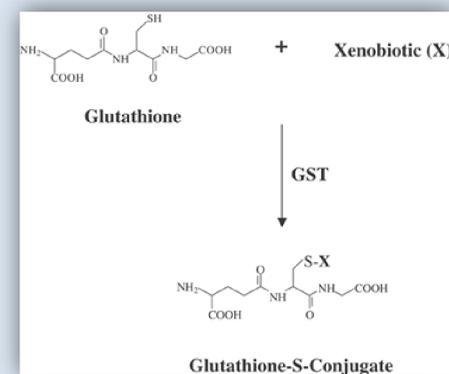
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Introduction

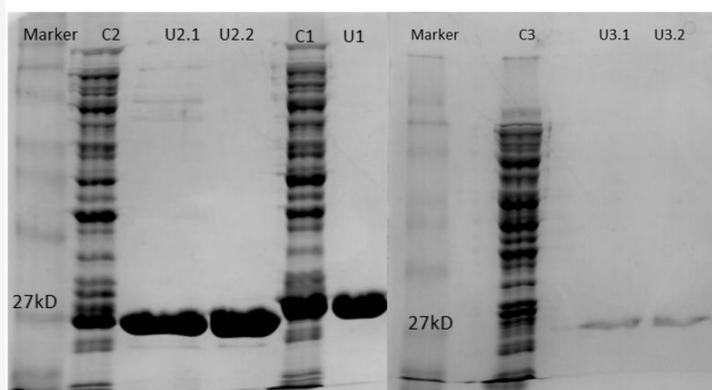
GSTs are multifunctional enzymes that catalyze the conjugation of glutathione (GSH) to reactive electrophiles. These electrophiles are diverse and include important endogenous compounds, as well as xenobiotic chemicals, therefore GSTs play an important role in stress tolerance and herbicide detoxification. GSTs are usually active as a dimer of 24–29 kDa subunits. Each monomer of dimeric GSTs contains a G-site, at the N-terminal, capable of binding the GSH substrate and an H-site, at the C-terminal, that has xenobiotic compound-binding capabilities. Different classes of herbicides such as triazines, thiocarbamates, chloroacetanilides, diphenylethers, and aryloxyphenoxypropionates can be metabolized by GSTs. Herbicide tolerance in plants is based primarily on the differential ability of plant species to detoxify a herbicide, with the formation of a herbicide-GSH conjugate in the resistant but not in the susceptible species. The plant-specific phi and tau GSTs are primarily responsible for herbicide detoxification, showing class specificity in substrate preference. In present work, we report the cloning, kinetic and structural characterization of three members of the GST family from *Cicer arietinum* leaves (CaGSTs).

Methods and materials

GSTs of *Cicer arietinum* were identified by in silico transcriptome analysis and their full length cDNAs with complete open reading frame were isolated using RT-PCR. Analysis of the cDNA clones showed that the deduced amino acid sequences share high homology with GSTs that belong to tau classes. The GST isoenzymes were expressed in *E. coli* and purified by affinity chromatography and their substrate specificity was determined towards nineteen different electrophilic substrates including herbicides. Comparative molecular modelling was used to identify key structural characteristics and to provide insights into the substrate specificity and the catalytic mechanism of these enzymes.



Results



- Purified recombinant CaGST-U1, CaGST-U2 and CaGST-U3 (Fig 1) were assayed as glutathione transferase, glutathione peroxidase, dehydroascorbate reductase and as thioltransferase and specific activities were determined using different substrates (Fig 2).

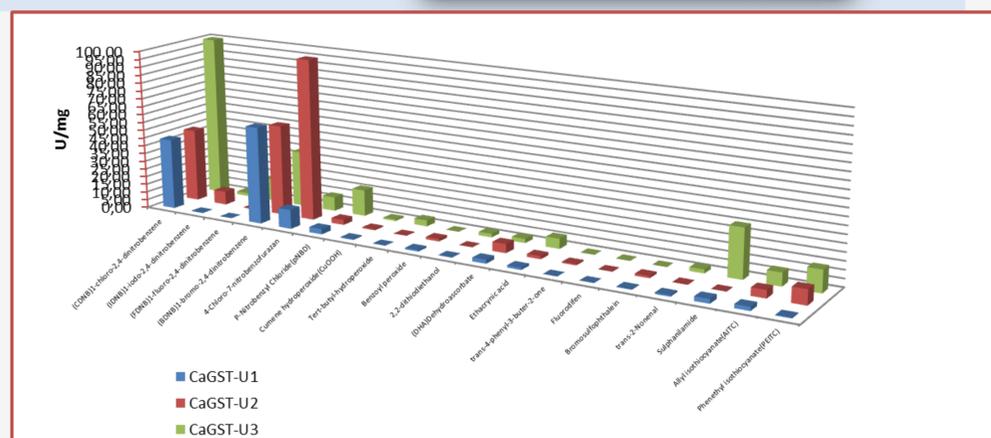


Fig 2: Substrate specificity for purified recombinant CaGST-U1, CaGST-U2, CaGST-U3, towards 19 different electrophilic substrates. Enzyme assays were carried out under standard conditions. Results represent the means of triplicate determinations.

Fig 1: SDS-PAGE analysis of purified CaGST-U1, CaGST-U2, CaGST-U3. Elution was carried out using GSH (10 mM). M protein markers, lane C1, C2, C3 crude extract, lanes U1-U3 eluted fractions from the affinity column.

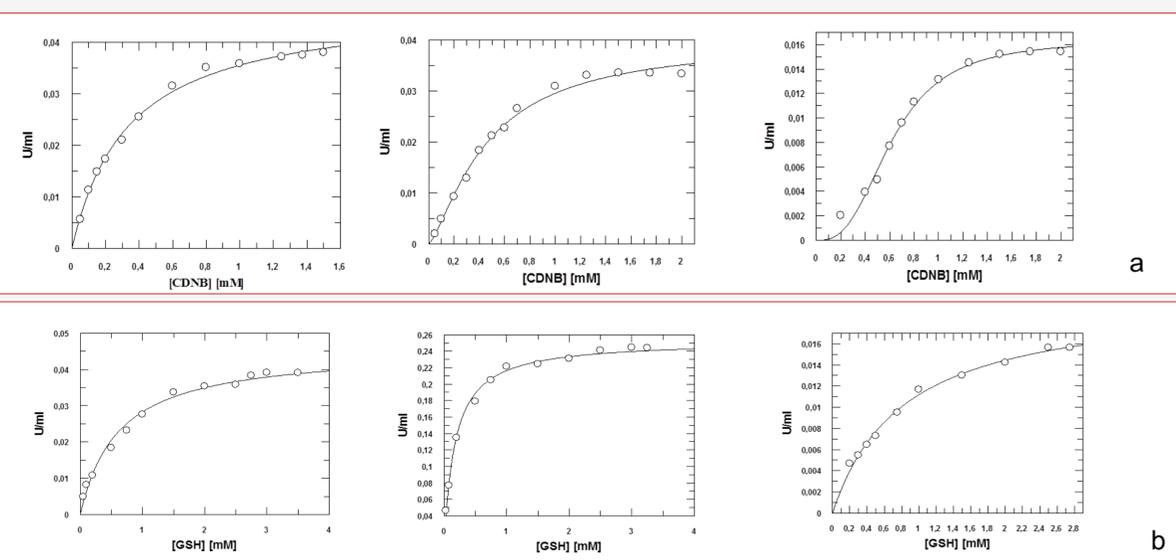


Fig 3: Kinetic analysis of CaGST-U1, CaGST-U2, CaGST-U3 using the CDNB as a variable substrate (a) and GSH at a fixed concentration. Kinetic analysis of CaGST-U1, CaGST-U2, CaGST-U3 using the GSH as variable substrate (b) and CDNB at a fixed concentration. Experiments were performed in triplicate, and lines were those calculated by least-squares regression analysis.

Table 1. Kinetic analysis of CaGSTs.

	K_m (mM) (CDNB)	K_{cat} (min^{-1}) (CDNB)	K_{cat}/K_m ($\text{mM}^{-1}\text{min}^{-1}$)(CDNB)
CaGST-U1	$0,3443 \pm 0,0184$	694	2015,7
	$S_{0,5}$ (mM)(CDNB)	K_{cat} (min^{-1}) (CDNB)	η_H (CDNB)
CaGST-U2	$0,3508 \pm 0,0611$	347	1,535
CaGST-U3	$0,4644 \pm 0,1687$	206	2,0904

	K_m (mM) (GSH)	K_{cat} (min^{-1}) (GSH)	K_{cat}/K_m ($\text{mM}^{-1}\text{min}^{-1}$)(GSH)
CaGST-U1	$0,6128 \pm 0,0731$	664	1083
CaGST-U2	$0,1707 \pm 0,0122$	154	902
CaGST-U3	$0,8221 \pm 0,0597$	206	250

- The kinetic parameters k_{cat} , K_m , $S_{0,5}$ and η_H were determined by steady-state kinetic analysis (Fig 3) and the results are listed in Table 1.

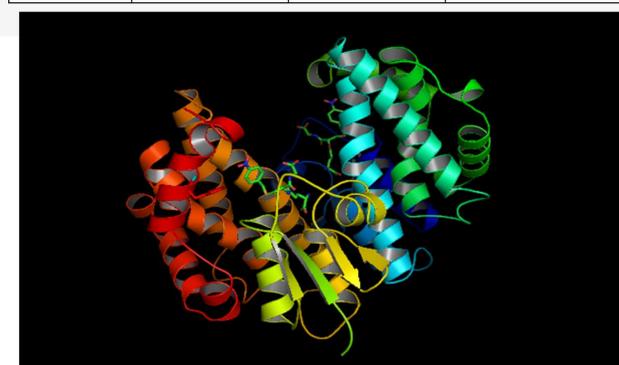


Fig. 4 :A Ribbon diagram of the CaGST-U1 protein model. The figure was created using PyMOL.

Discussion

- The results showed that the glutathione transferases from *C. arietinum* (CaGSTs) catalyze a broad range of reactions and exhibit quite varied substrate specificity.
- CaGST-U1 obeys Michaelis - Menten hyperbolic equation using CDNB and GSH as a variable substrate. Steady-state kinetic analysis of CaGST-U2 and CaGST-U3 showed that when CDNB was used as a variable substrate with GSH at fixed concentration, a sigmoid substrate dependence was observed.
- Molecular modelling showed that the isoenzymes share the same structural organization with other plant cytosolic GSTs and some differences at the level of C-terminal domain (Fig 4). In particular, major variations were identified in helices H4 and H9 that affect xenobiotic substrate recognition and catalytic mechanism.