

# A workflow for the orthogonal identification of biotransformation products by HILIC-QTOFMS



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CEST 2015

Determine the degradation rate of emerging contaminants (pharmaceuticals, illicit drugs) in the aquatic environment under aerobic conditions

Study the occurrence and formation of transformation products

Develop an integrated workflow for suspect and non-target screening for the identification of biotransformation products

Study the use of HILIC as a complementary technique for the identified TPs

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Further investigation for suspect and unknown TPs which may be eluted better in HILIC conditions







Samples of activated sludge from active bioreactors and effluent waste water from the WWTP of Athens (Psitalia)



Batch reactors seeded with activated sludge under aerobic conditions in room temperature



Spike in a concentration of 2 mg/L



Sampling immediately after spiking and after several time intervals (depending on the compound)

Sauth 1

Filtration through glass fiber syringe and then through 0.2  $\mu m$  RC filter

LC-HRMS (qTOF) both RP and  $\underline{\text{HILIC}}$ 



#### Instrumentation

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UltiMate 3000™ RSLC (Dionex) UHPLC System Column: Thermo Acclaim RSLC C18, 2.2µm 120 Å, 2.1x100 mm Mobile Phase & gradient program: Bruker Pesticide screener RP Inj. Volume: 5 µL Column: Waters BEH Amide Acquity 1.7 µm, 2.1×100 mm Inj. Volume: 5 µL HILIC

> Bruker Maxis Impact™ Quadrupole-Time-of-Flight



#### Levels of identification confidence

Identification confidence



#### Example

Minimum data requirements

H <sub>3</sub> C <sub>s</sub>	Level 1:	<b>Confirmed structure</b> by reference standard	MS, MS <sup>2</sup> , RT, Reference Std.	
	Level 2:	<i>Probable structure</i> a) by library spectrum match b) by diagnostic evidence	MS, MS <sup>2</sup> , Library MS <sup>2</sup> MS, MS <sup>2</sup> , Exp. data	
	Level 3:	<i>Tentative candidate(s)</i> structure, substituent, class	MS, MS <sup>2</sup> , Exp. data	
C <sub>6</sub> H <sub>5</sub> N <sub>3</sub> O <sub>4</sub>	Level 4:	Unequivocal molecular formula	MS isotope/adduct	
192.0757	Level 5:	Exact mass of interest	MS	

Schymanski et al. Environmental Science and Technology (2014) 48(4):2097



**Compilation** of suspect list: Metabolite prediction tools+ literature

**MassBank** 

MetFrag

**MetFUS** 

Screening all the time interval chromatograms both in RP and HILIC (+ESI/-ESI)  $\implies$  XIC

**Evaluation of candidates** (Tentative): meet the set criteria, absence in the blank, chromatographic retention time plausibility (time trend)

Acquire MS/MS spectra with inclusion list both in RP and HILIC: Interpretation of the fragmentation pathway

**Confirmation**, if it's possible with reference standard (RT and MS/MS spectra)



#### Compilation of the suspect list

EAWAG-BBD Pathway Prediction System

http://eawagbbd.ethz.ch/predict/ Rule based

Microbial catabolic reactions

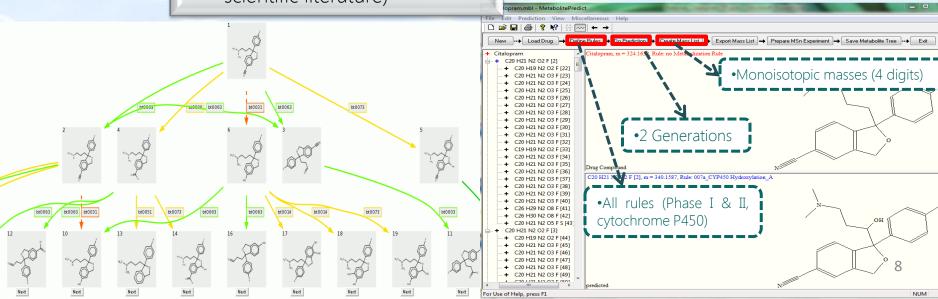
(reactions found in the EAWAG-BBD database/

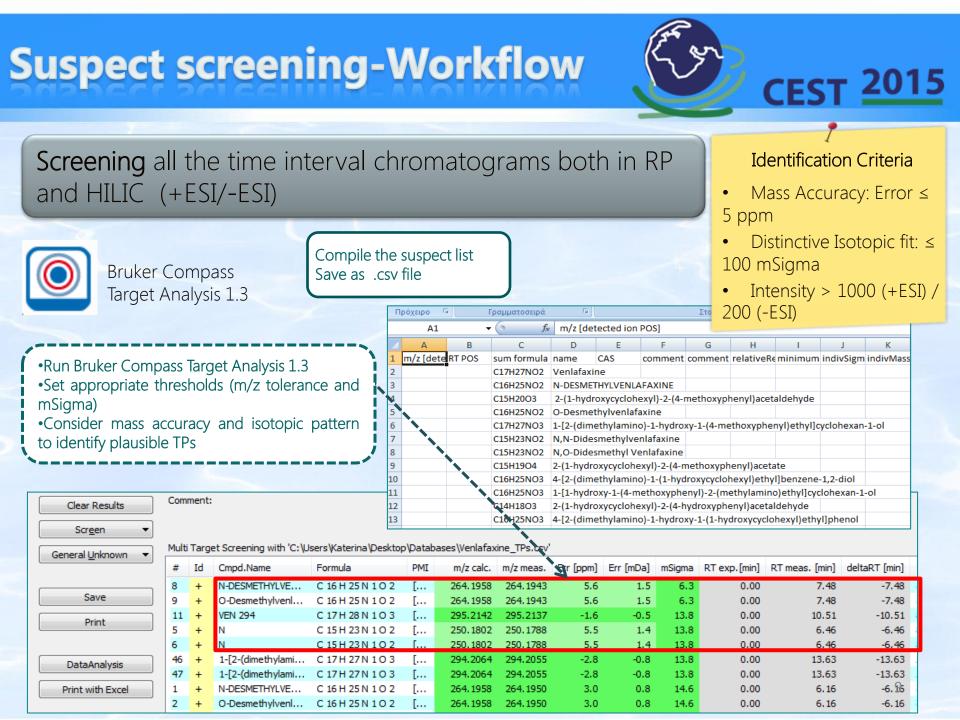
scientific literature)

Metabolite Predict

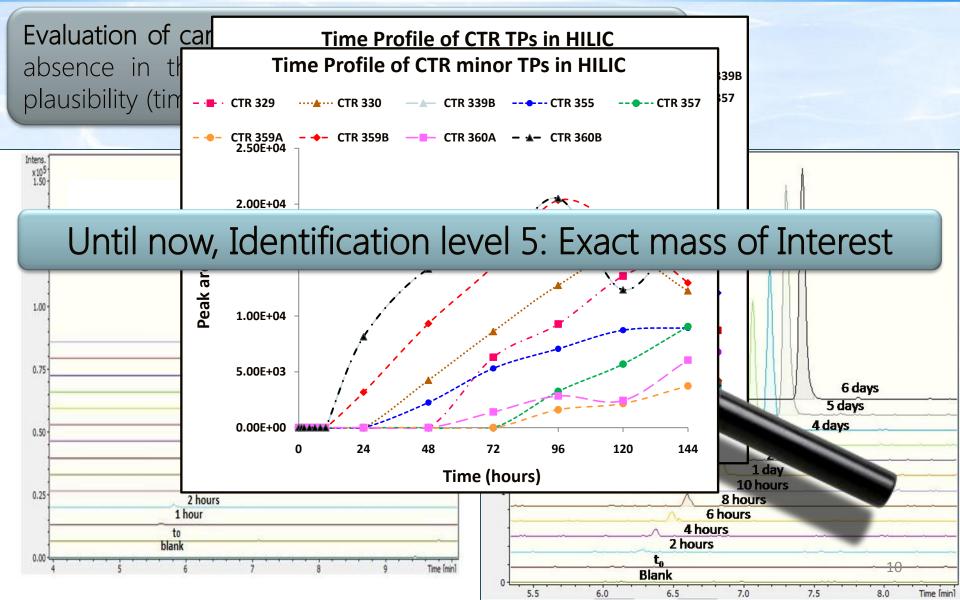
Metabolomic software Rule based (Phase I & II metabolism,

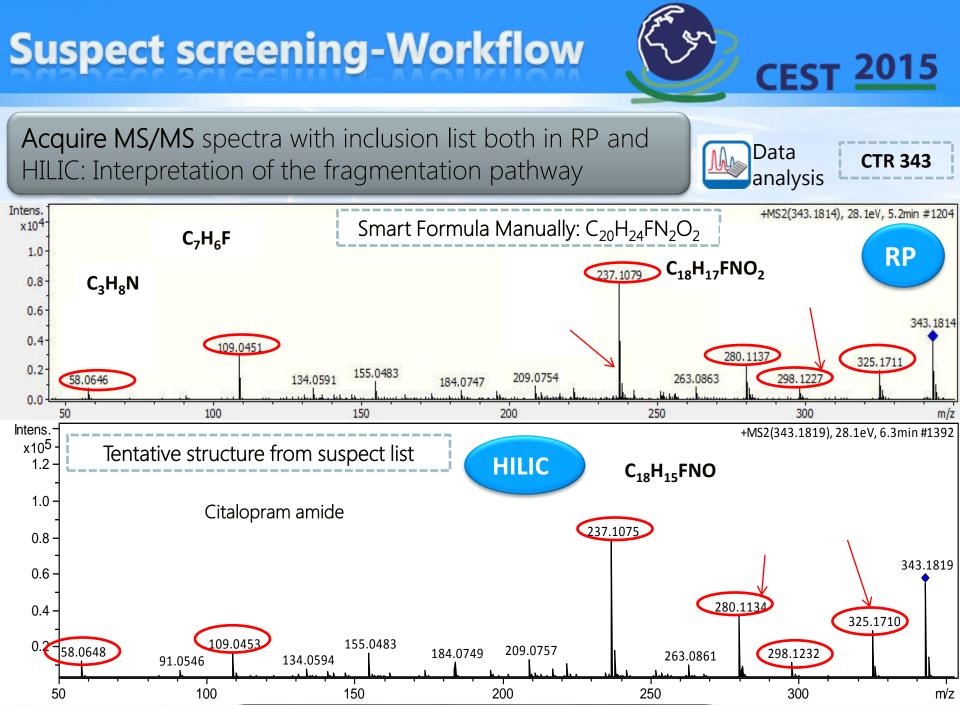
cytochrome P450)

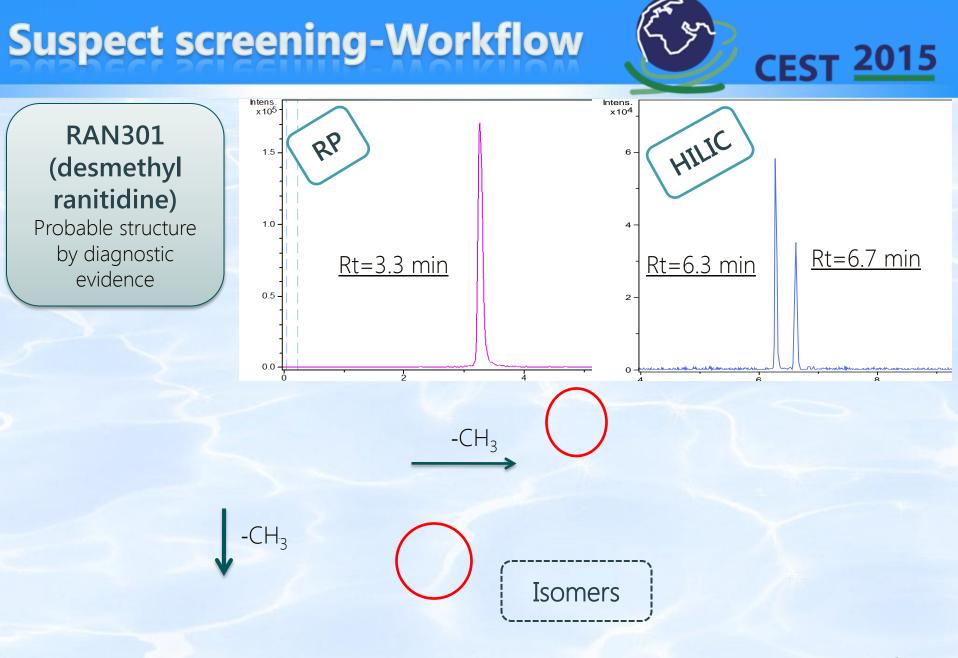




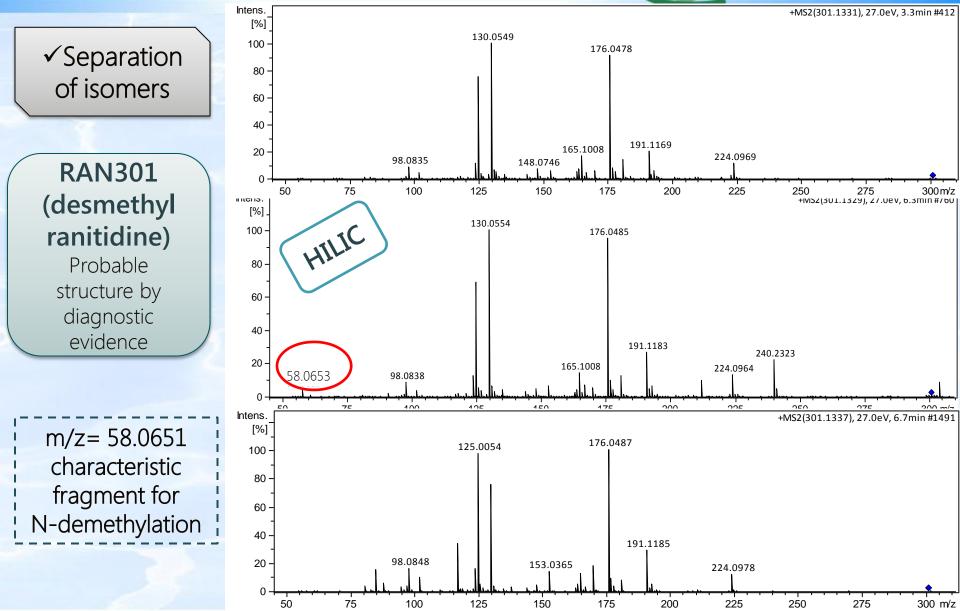


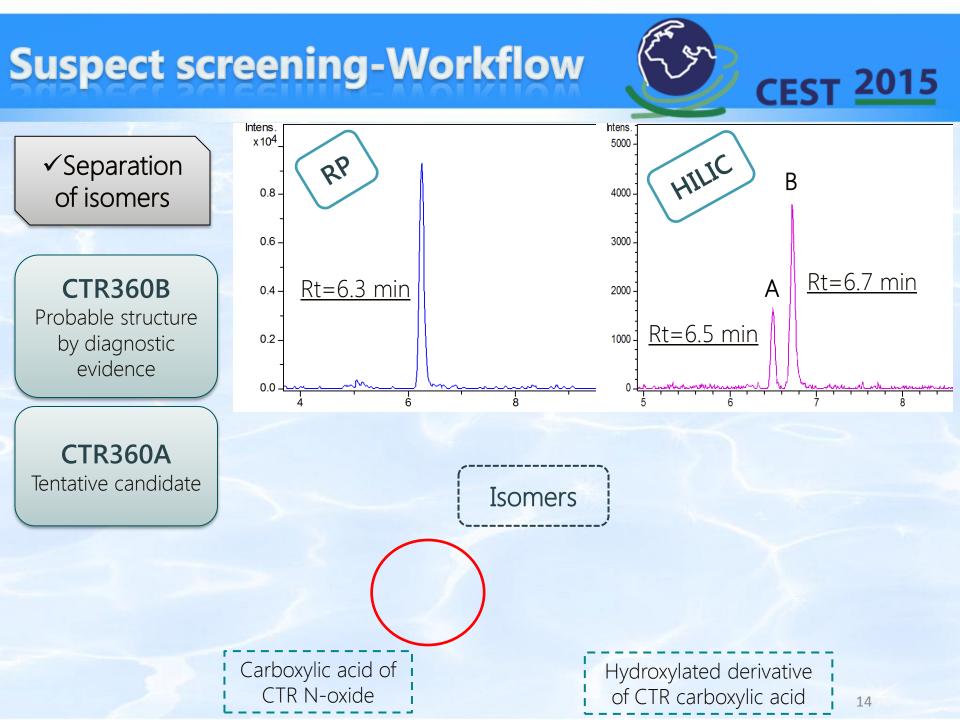


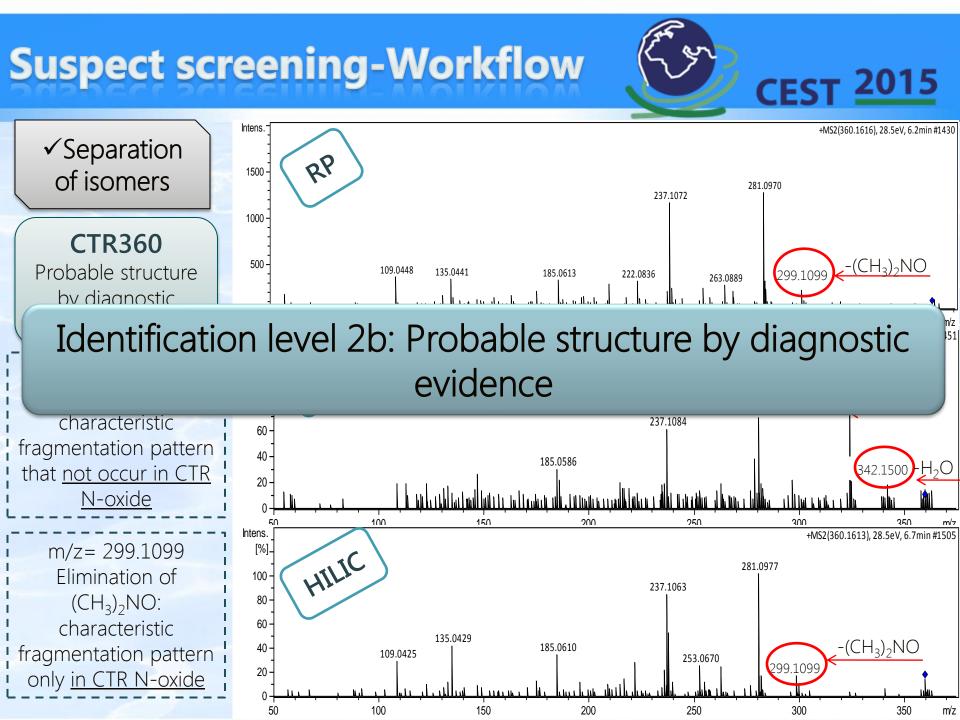


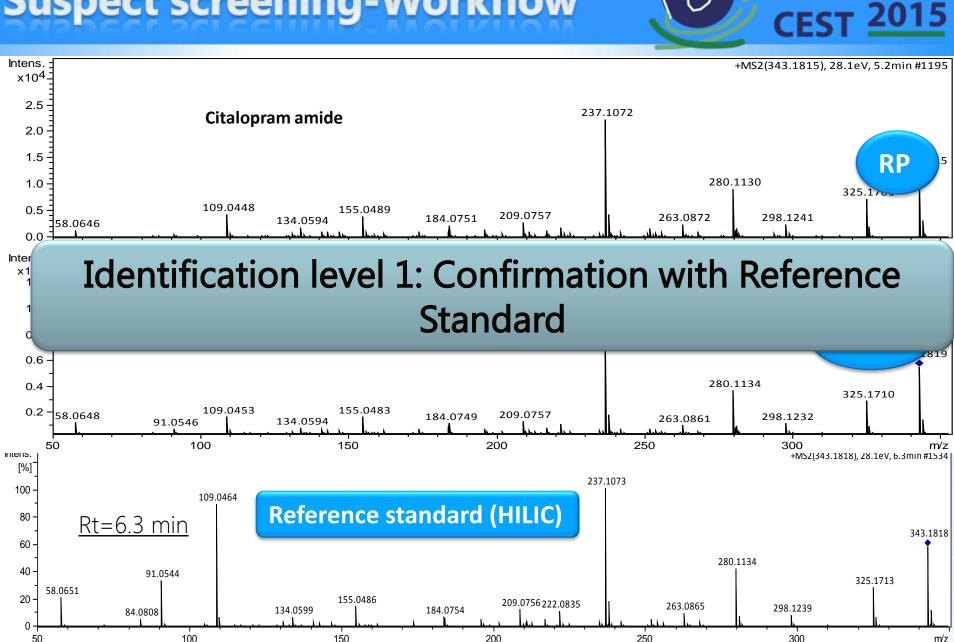


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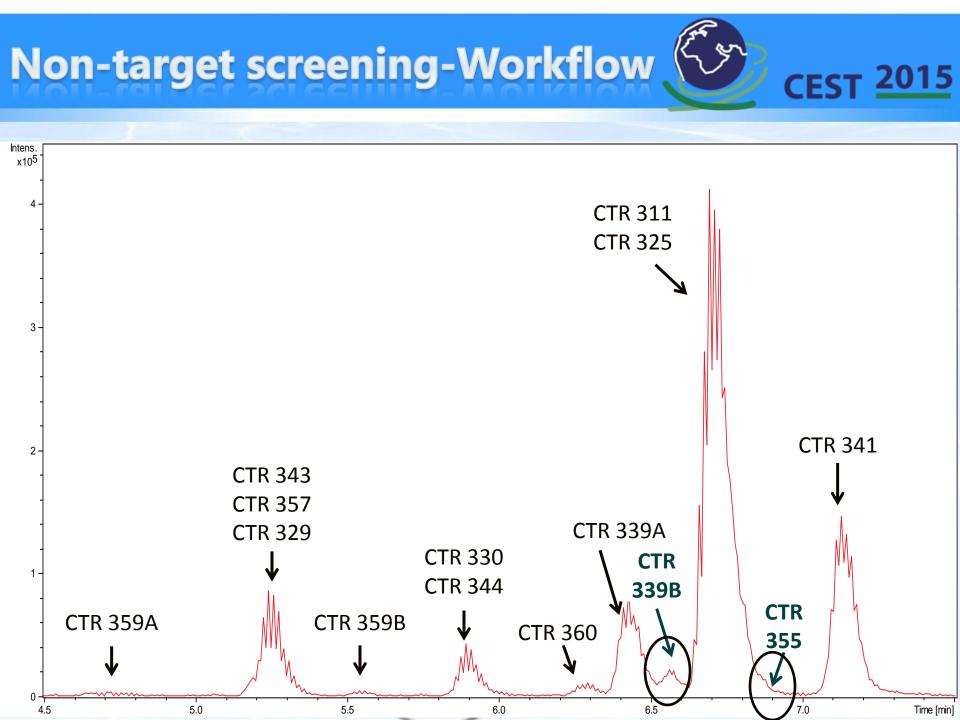




Non-target screening-Workflow

Subtraction of the background (Bruker's MetaboliteDetect 2.0) **Background Subtraction Parameters** Generation of a peak list of unknown Difference eXpose mode, Ratio: 3 Detect -Mass spectrum Parameters Treat further as "suspect peaks" Metabolite Int threshold: 30% Detect Max No. of peaks: 20 Sample Chromatogram detected expected Generation of peak list Intens.: Sample x106 RT-S[min] Gp. No. Chrom. m/z-det. M2/M2 Comment 1.5 Base Peak Base Peak Ch 1.0 R BPC Int. Base Peak Ch 0.5 TIC Total Ion Curr 0.0 D EIC Extracted Ion **(** 12 ó ż Ġ. ģ 10 14 16 18 Time [min] 1\* 1.3 107.9681 Det. Trace 1 (yes) Reference Chromatogram 2 1 1.3 123,9411 (yes) Det. Trace 2 Intens.-3 1 1.3 139.9146 (yes) Det. Trace 3 **Blank Sample** x106 2\* 3.4 216.1231 (yes) Det. Trace 4 4.2 207.1489 5 3a1\* (yes) Det. Trace 5 4.2 266.1975 6 3b1 (yes) Det. Trace 6 4\* 4.5 86.0973 Det. Trace 7 (yes) **(** 2 6 8 10 12 14 16 18 Time [min] 5\* 5.2 86.0963 Det. Trace 8 (yes) g 6\* 6.7 219.1491 Det. Trace 9 (yes) Difference Chromatogram 10 7\* 9.4 652.2653 Det. Trace 10 (yes) Intens. **Background** x106 7 11 ? 694.2812 Det. Trace 11 (ves) Subtraction 0.75 12 8\* 10.6 910.5027 (ves) Det. Trace 12 0.50 0.25 0.00 **(** 14 Time [min] 6 10 12 2 4 Ś.

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#### Results



### **CEST** 2015

			Appearance of max peak area	Molecular Formula	Mass [M+H] <sup>+</sup>	ТР	PC`
↗↘ 1 MEGX	1	7∖	7d	C12H18N2O	207.1504	LDC207	Lidocaine
↗↘ 1 Lidocaine-N-oxide	1	<i>7</i> ∖	2d	C14H22N2O2	251.1756	LDC251	
<b>⊅</b> ∿ 2b	21	<i>7</i> ∖	1d	C13H18N2O	219.1485	LDC219	
$\nearrow \rightarrow 4$	4	$\nearrow$	7d	C13H16N2O2	233.1282	LDC233	
$\nearrow$ 4	4	$\nearrow$	9d	C14H20N2O3	265.1538	LDC265	
↗ 1 Desmethyl CTR	1	7	6 d	C19H19FN2O	311.1554	CTR311	Citalopram
↗ 2b Desmethyl CTR amide	2b	7	6 d	C19H21FN2O2	329.1660	CTR329	
↗ 2b Desmethyl CTR carboxylic acid	2b	7	6 d	C19H20FNO3	330.1500	CTR330	
↗ 1 3-Oxo-CTR	1	7	6 d	C20H19FN2O2	339.1503	CTR339A	
↗ 2b Methylated derivative of CTR	2k	7	6 d	C21H23FN2O	339.1880	CTR339B	
↗↘ 2a CTR-N-oxide	28	$\nearrow$	3 d	C20H21FN2O2	341.1660	CTR341	
↗ 1 CTR amide	1	7	6 d	C20H23FN2O2	343.1816	CTR343	
↗ 1 CTR carboxylic acid	1	7	5 d	C20H22FNO3	344.1656	CTR344	
↗ 3 Hydroxylated derivative of 3-Oxo-CTR	3	7	6 d	C20H19FN2O3	355.1457	CTR355	
↗ 2b Amide of 3-Oxo-CTR	2k	7	6 d	C20H21FN2O3	357.1609	CTR357	
↗ 3 Hydroxylated derivative of CTR amide	3	7	6 d	C20H23FN2O3	9A 250 1765	CTR359A	
↗> 2b Amide of CTR-N-oxide	21	$\mathbb{Z}$	4 d		CTR359B		
↗ 3 Hydroxylated derivative of CTR carb.acid	3	7	6 d	C20H22FNO4	CTR360A 360 1606		
↗ 2b Carboxylic acid of CTR-N-oxide	2k	7			500.1000	CTR360B	
↗ 1 Guanylurea	1	7	2d	C2H6N4O	103.0614	MTF103	Mt
<ul> <li>2b Desmethyl CTR carboxylic aci</li> <li>1 3-Oxo-CTR</li> <li>2b Methylated derivative of CTR</li> <li>2a CTR-N-oxide</li> <li>1 CTR amide</li> <li>1 CTR carboxylic acid</li> <li>1 CTR carboxylic acid</li> <li>3 Hydroxylated derivative of 3-0</li> <li>2b Amide of 3-Oxo-CTR</li> <li>3 Hydroxylated derivative of CT</li> <li>2b Amide of CTR-N-oxide</li> <li>3 Hydroxylated derivative of CT</li> <li>3 Hydroxylated derivative of CT</li> <li>2b Amide of CTR-N-oxide</li> <li>3 Hydroxylated derivative of CT</li> <li>2b Amide of CTR-N-oxide</li> <li>3 Hydroxylated derivative of CT</li> <li>2b Amide of CTR-N-oxide</li> <li>2b Carboxylic acid of CTR-N-oxide</li> </ul>	2b 1 2k 2a 1 1 3 2k 3 2k 3 2k	アスアンティンション	6 d 6 d 3 d 6 d 5 d 6 d 6 d 6 d 4 d 6 d	C19H20FNO3 C20H19FN2O2 C21H23FN2O C20H21FN2O2 C20H23FN2O2 C20H22FNO3 C20H19FN2O3 C20H21FN2O3 C20H21FN2O3	330.1500 339.1503 339.1880 341.1660 343.1816 344.1656 355.1457 357.1609 359.1765 360.1606	CTR330 CTR339A CTR339B CTR341 CTR343 CTR344 CTR355 CTR357 CTR359A CTR359B CTR360A CTR360B	Mtf Citalopram

#### Results



PC	ТР	Mass [M+H] <sup>+</sup>	Molecular Formula	Appearance of max peak area	Time trend	Id. Level	Name
	RAN286A		C11H15N3O4S	1d	75	4	-
	RAN286B	286.0856		2d	$\nearrow$	4	-
	RAN286C			10h	ブン	4	-
	RAN301A	201 1 2 2 0	C12H20N4O3S	10h	75	2b	
	RAN301B	301.1329				2b	desmethyl RAN
dine	RAN302A	202 0805	C11H15N3O5S	ld	7	3	
Ranitidine	RAN302B	502.0605		1d	$\nearrow$	3	
	RAN316A (2peaks)	316.1331		3d	7	3	
	RAN316B			3d	7	3	
	RAN317 (2peaks)	317.1278	C12H20N4O4S	3d	7	3	Desmethyl RAN-S-oxide
	RAN331A (2peaks)	331.1435	C13H22N4O4S	1d	75	1	RAN-S-oxide
	RAN331B			1d	7∖	1	RAN-N-oxide
atin	ATR515	515.2341	C31H31FN2O4	2d	7	2b	$\beta$ -oxidation product of ATR
'asta	ATR471	471.2078	C29H27FN2O3	2d	7	2b	$\beta$ -oxidation product of ATR515
Atorvastatin	ATR487A ATR487B 487.2028		3 C29H27FN2O4	2d	7	3	Monohydroxylation of ATR471
				2d	7	3	





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5 pharmaceuticals (metformin, ranitidine, lidocaine, citalopram and atorvastatine) were investigated for their TPs under aerobic biodegradation experiments and 34 TPs were identified, in total.



Nine of them were confirmed by a reference standard and the majority of the rest reached Id. Level 2: probable structure by diagnostic evidence.



HILIC was presented to be fit for the orthogonal identification of TPs and suitable for screening of more polar metabolites



HILIC provided better peak shapes and greater intensities in some of the identified TPs.



HILIC also, permitted the identification and characterisation of isomeric TPs, due to the better detection sensitivity that provided clearer spectra for interpretation.





MANAGING



#### TREMEP

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AUTHORITY

# Image: A constraint of the const

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#### Thank you for your attention



http://trams.chem.uoa.gr/ http://tremepol.chem.uoa.gr/